

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

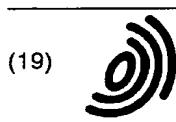
Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,
please do not report the images to the
Image Problem Mailbox.**



(19)

Europäisches Patentamt
European Patent Office
Office européen des brevets



(11)

EP 1 134 232 A1

(12)

EUROPEAN PATENT APPLICATION

(43) Date of publication:
19.09.2001 Bulletin 2001/38

(51) Int Cl.7: C08B 37/00, C08B 11/20,
A61L 15/28, A61L 15/00

(21) Application number: 00200965.2

(22) Date of filing: 16.03.2000

(84) Designated Contracting States:
AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU
MC NL PT SE

Designated-Extension States:
AL LT LV MK RO SI

- Porro, Fabrizio
80055 Portici (NA) (IT)
- Nicolais, Luigi
80056 Ercolano (NA) (IT)
- Sannino, Alessandro
80055 Portici (NA) (IT)

(71) Applicant: SCA Hygiene Products AB
405 03 Göteborg (SE)

(74) Representative: Jorritsma, Ruurd et al
Nederlandsch Octrooilbureau
Scheveningseweg 82
P.O. Box 29720
2502 LS Den Haag (NL)

(54) Polysaccharide-based superabsorbent film

(57) A superabsorbent polysaccharide can be obtained by crosslinking a polysaccharide or derivative thereof with at least 1 % by weight of a flexible spacer having a chain length of at least 9 chain atoms and hav-

ing terminal activated coupling groups. The flexible spacer may comprise a polyalkyleneglycol with a molecular weight from about 400 to 10,000. The coupling groups may be provided by divinyl sulphone units.

EP 1 134 232 A1

Description

Field of the Invention

5 [0001] The present invention relates to flexible superabsorbent films based on polysaccharides such as cellulose and derivatives thereof, and to a process for producing such films.

Background

10 [0002] Superabsorbent materials for use in hygiene products, which are based on polysaccharides such as cellulose and starch, have recently become widely known in the art, for example in WO 98/27117. The absorbing capacity of such materials can be increased by crosslinking the polymers, e.g. by using epichlorohydrin, diglycidyl ethers, divinyl sulphone or other commonly known crosslinkers capable of reacting with hydroxyl groups, or by using carboxylated polysaccharides and crosslinkers capable of reacting with carboxyl groups, such as divalent metals. However, there is a demand for thinner absorbent products, which implies that superabsorbent materials have to be found which have further increased absorbing capacity and have increased flexibility.

15 [0003] WO 97/19116 describes superabsorbent acrylic polymers which are crosslinked by polymerisation of acrylic acid in the presence of a combination of trimethylolpropane triacrylate or triallylamine, polyethyleneglycol mono(meth)acrylate monoalkyl ether and polyethyleneglycol mono(meth)acrylate monoalkyl ether.

20 [0004] WO 97/31971 discloses similar, foamed superabsorbent acrylic polymers which are crosslinked e.g. with trimethylolpropane triacrylate, to which internal or external plasticisers (e.g. glycerol or acrylic esters) may be added to increase flexibility of the foam.

Description of the Invention

25 [0005] It has been found that thin superabsorbent polysaccharides with high absorption capacity and sufficient flexibility can be obtained by crosslinking the polysaccharides with flexible spacers such as polyalkyleneglycols, having terminal activated groups. The products and the process of producing them are defined in the appended claims.

30 [0006] The polysaccharides to be used according to the present invention are in particular α -glucans like starch, amylose and amylopectin, β -glucans like cellulose, galactomannans like guar gum (guaran) and locust bean gum, glucomannans including e.g. xanthan gum, fructans, (arabino)xylyans and galactans, as well as derivatives such as carboxymethyl, alkyl, hydroxyethyl and hydroxypropyl derivatives of such polysaccharides. Cellulose and cellulose derivatives are preferred for practical reasons. Combinations of such polysaccharides, or combinations with other polymers such as polyacrylates, polyvinyl alcohol etc. can also be used. The chain length of the polysaccharides is important, although there is no critical minimum for the molecular weight. In general, polysaccharides having a molecular weight of more than 25,000 are preferred.

35 [0007] The polysaccharides to be used according to the present invention may also be carboxymethylated or carboxyethylated, especially in the case of cellulose. Other carboxy-alkylated polysaccharides include the half esters obtained from cyclic anhydrides such as succinic and maleic anhydride, and addition products of maleic half esters to which sulphite has been added. The degree of carboxyalkylation is preferably between 0 and 1.5, in particular between 0.1 and 1.0 carboxyalkyl groups per monosaccharide unit. The carboxyl derivatives may be in their acid or in salt form. Combinations of carboxylated polysaccharides such as CMC (carboxymethyl cellulose) and hydroxyalkylated polysaccharides (e.g. hydroxyethyl cellulose, HEC) are especially useful, whether as mixtures of different derivatives (e.g. HEC and CMC, or HEC and carboxymethyl starch, or HEC and methyl cellulose) or as multiply derivatised single compounds (e.g. sodium carboxymethyl-hydroxyethyl cellulose, CMHEC).

40 [0008] The polyalkyleneglycols to be used as spacers may for example be polyethyleneglycol (PEG), polypropyleneglycol (PPG) and the like. Other hydrophilic or hydrophobic spacers may also be used, as long as they are flexible, i.e. contain no or only a few double bonds or cyclic structures; examples are polyalkylene (as in decamethylene diisocyanate), polyhydroxyalkylene, polyalkylene succinate, polylactide, etc, with chain lengths from about 9 to about 750 chain atoms. The chain length of the polyalkyleneglycols may vary from e.g. 3 units (MW about 150 Da) up to e.g. 250 (MW about 11,000). Molecular weights from about 1000 to about 8000 are preferred. The relative amount of polyalkyleneglycol with respect to the polysaccharide may vary from about 1/200 to about 1/1, especially from about 1/50 to about 1/1.5 (weight ratios), depending on the required thickness and the required flexibility of the product.

45 [0009] The terminal activated groups are preferably vinyl groups activated by carbonyl or sulphonyl functions, for example acryloyl groups (-CO-CR=CHR), maleoyl groups (-CO-CH=CH-COOH) or vinylsulphonyl groups (-SO₂-CR=CHR), in which each R may be hydrogen (preferred), methyl or other alkyl. Such groups may be directly attached to the polyalkyleneglycol, e.g. as (sulphonate) esters, or through alkylene or phenylene groups. Particularly advantageous is the coupling product of a polyalkyleneglycol with divinyl sulphone on either side of the polyglycol. Other

EP 1 134 232 A1

terminal crosslinkers include (activated) halomethyl, activated hydroxymethyl, activated formyl, epoxy, isocyanate, and the like. Examples of such coupling agents (other than divinyl sulphone) are maleic anhydride, dichloroacetone, 1,3-dichloro-2-propanol, dimethylolurea, dimethylolimidazolidone, diepoxides such as bisepoxybutane or bis(glycidyl ether), epichlorohydrin, diisocyanates, bis(2-hydroxyethyl) sulphone, formaldehyde, glyoxal. The weight ratio between terminal crosslinker (such as divinyl sulphone) and spacer (such as polyalkylene glycol) can be between about 1/1 to about 100/1, especially between about 1.5/1 and 30/1. The weight ratio between crosslinker and polysaccharide may vary from e.g. 1/1 to 1/50, especially from 1/1.5 to 1/10.

[0010] The production of the superabsorbent films according to the invention can be divided in three steps: (1) mixing of reactants and other compounds, (2) reaction and washing stage, and (3) desiccation. As to step (1), the components involved in the reaction can be divided in different classes: (a) components of the base structure of the network, i.e. the polysaccharides, e.g. carboxymethyl cellulose sodium salt (CMCNa) and/or hydroxyethyl cellulose (HEC); (b) crosslinkers, e.g. divinyl sulphone (DVS); (c) spacers, e.g. polyethylene glycol (PEG); (d) catalysts, e.g. KOH; and solvents, e.g. water. In step (2), the reactants are allowed to react for a sufficient time to allow the production of a crosslinked gel. Preferably, the polyalkylene glycol and the reagent introducing the terminal double bonds are reacted first, followed by reaction with the polysaccharide, preferably in the presence of an alkaline catalyst. The crosslinking reaction can be performed at varying temperatures e.g. from about 5°C to about 40°C, for about 1 hour to about 2 days, preferably from 5-24 hours. After the crosslinking, the unreacted reagents can be removed by washing in distilled water, if desired, followed by drying. The crosslinked product can also be directly dried without a washing step.

[0011] The superabsorbent products according to the invention are flexible films with thicknesses between 10 and 500 µm and having absorption capacities between about 15 and 30 g of synthetic urine (300 mM urea, 60 mM KC1, 130 mM NaCl, 2.0 mM CaSO₄, 3.5 mM MgSO₄, 29 mM KH₂PO₄, 5.3 mM Na₂HPO₄, 1 mg/l Triton X-100 in deionised water) per g of product. They can be used in absorbent articles, such as diapers, incontinence guards, sanitary napkins, and the like. They can also be used in tissue papers including kitchen towels, napkins, industrial wipes and the like.

25 **Examples**

[0012] Materials: Divinyl sulphone (DVS), polyethyleneglycol (PEG) with various molecular weights (400, 4600, 10,000), hydroxyethyl cellulose (HEC, MW 250,000) and carboxymethyl cellulose (CMCNa, MW 700,000) were obtained from Aldrich Chimica, Milano, IT.

30 [0013] The amounts of reagents are given in the tables, per 150 ml of distilled water. DVS was dissolved in distilled water to a concentration of 40 mmol/l. PEG was then added to the DVS solution. After dissolution of the PEG the CMCNa and HEC were added in powder form and dissolved up to a concentration of about 2% (see tables). Best results were obtained by first dissolving HEC and then slowly admixing CMCNa. Mixing was continued at 25°C until a clear solution was obtained. After complete mixing, 1M of aqueous KOH was dissolved into the mixture up to the desired concentration. After another two minutes of stirring, the reaction mixture was spread on a teflon sheet with a Gardner knife in order to obtain a film with a controlled thickness. The film was allowed to crosslink at ambient temperature for between 5 and 24 hours (best results after 10-14 hours). Higher temperatures did not increase the crosslinking rate, and resulted in decreased viscosity. A thin, partially swollen gel film was obtained.

40 [0014] From this point on, two different procedures were followed. According to the first procedure, the teflon sheet with the partly swollen film was then put in a jar containing distilled water. As soon as the film started to swell further, the teflon sheet was removed. During swelling, water mixture containing residual KOH, unreacted DVS and other impurities was continuously removed from the bottom of the jar, while fresh distilled water was added. After equilibrium swelling occurred, the teflon sheet was again positioned under the film, water around the film was removed and the film was dried under atmospheric conditions.

45 [0015] According to the second procedure, the washing (addition and removal of water) was omitted and the swelling film was maintained on the teflon sheet for 5-24 hours and then dried under atmospheric conditions.

[0016] As an alternative to drying under atmospheric conditions (for about 6-20 days), desiccation was performed in an oven at 50-100 °C, with best results being obtained at 60-80°C, for 1-24 hours.

50 Table 1

Hydrogel synthesis mixture with PEG 400			
Molar ratio [PEG]/[DVS] = 1/30; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
Water	150	8330 + 280 ¹	94.54

55 ¹ the water of the KOH solution

EP 1 134 232 A1

Table 1 (continued)

Hydrogel synthesis mixture with PEG 400			
Molar ratio [PEG]/[DVS] = 1/30; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
CMCNa	2.25	3.21×10^{-3}	1.42
HEC	0.75	3.00×10^{-3}	0.47
KOH, 1M in water	5.28	KOH: 5.00	3.33
DVS	0.35	2.96	0.22
PEG 400	0.04	0.100	0.03

15

Table 2

Hydrogel synthesis mixture with PEG 400			
Molar ratio [PEG]/[DVS] = 1/90; molar ratio [PEG]/[cellulose] = 11/1			
Reagent	grams	mmoles	% by weight
Water	150	$8330 + 280^1$	94.33
CMCNa	2.25	3.21×10^{-3}	1.41
HEC	0.75	3.00×10^{-3}	0.47
KOH, 1M in water	5.28	KOH: 5.00	3.32
DVS	0.71	6.01	0.45
PEG 400	0.027	0.0675	0.02

¹the water of the KOH solution

30

Table 3

Hydrogel synthesis mixture with PEG 400			
Molar ratio [PEG]/[DVS] = 1/60; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
Water	150	$8330 + 280^1$	94.32
CMCNa	2.25	3.21×10^{-3}	1.41
HEC	0.75	3.00×10^{-3}	0.47
KOH, 1M in water	5.28	KOH: 5.00	3.32
DVS	0.71	6.01	0.45
PEG 400	0.04	0.100	0.03

¹the water of the KOH solution

50

Table 4

Hydrogel synthesis mixture with PEG 400			
Molar ratio [PEG]/[DVS] = 1/10; molar ratio [PEG]/[cellulose] = 96/1			
Reagent	grams	mmoles	% by weight
Water	150	$8330 + 280^1$	94.20
CMCNa	2.25	3.21×10^{-3}	1.41

¹the water of the KOH solution

EP 1 134 232 A1

Table 4 (continued)

Hydrogel synthesis mixture with PEG 400			
Molar ratio [PEG]/[DVS] = 1/10; molar ratio [PEG]/[cellulose] = 96/1			
Reagent	grams	mmoles	% by weight
HEC	0.75	3.00×10^{-3}	0.47
KOH, 1M in water	5.28	KOH: 5.00	3.32
DVS	0.71	6.01	0.45
PEG 400	0.24	0.600	0.15

Table 5

Hydrogel synthesis mixture with PEG 400			
Molar ratio [PEG]/[DVS] = 1/200; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
Water	150	$8330 + 280^1$	93.36
CMCNa	2.25	3.21×10^{-3}	1.40
HEC	0.75	3.00×10^{-3}	0.47
KOH, 1M in water	5.28	KOH: 5.00	3.29
DVS	2.35	19.9	1.46
PEG 400	0.04	0.100	0.03

¹ the water of the KOH solution

Table 6

Hydrogel synthesis mixture with PEG 400			
Molar ratio [PEG]/[DVS] = 1/100; molar ratio [PEG]/[cellulose] = 32/1			
Reagent	grams	mmoles	% by weight
Water	150	$8330 + 280^1$	93.34
CMCNa	2.25	3.21×10^{-3}	1.40
HEC	0.75	3.00×10^{-3}	0.47
KOH, 1M in water	5.28	KOH: 5.00	3.29
DVS	2.35	19.9	1.46
PEG 400	0.08	0.200	0.05

¹ the water of the KOH solution

Table 7

Hydrogel synthesis mixture with PEG 4600			
Molar ratio [PEG]/[DVS] = 1/30; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
Water	150	$8330 + 280^1$	94.29
CMCNa	2.25	3.21×10^{-3}	1.41
HEC	0.75	3.00×10^{-3}	0.47

¹ the water of the KOH solution

EP 1 134 232 A1

Table 7 (continued)

Hydrogel synthesis mixture with PEG 4600			
Molar ratio [PEG]/[DVS] = 1/30; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
KOH, 1M in water	5.28	KOH: 5.00	3.32
DVS	0.35	2.96	0.22
PEG 400	0.46	0.100	0.29

Table 8

Hydrogel synthesis mixture with PEG 4600			
Molar ratio [PEG]/[DVS] = 1/60; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
Water	150	8330+280 ¹	94.07
CMCNa	2.25	3.21 * 10 ⁻³	1.41
HEC	0.75	3.00 * 10 ⁻³	0.47
KOH, 1M in water	5.28	KOH: 5.00	3.31
DVS	0.71	6.01	0.45
PEG 400	0.46	0.100	0.29

¹ the water of the KOH solution

Table 9

Hydrogel synthesis mixture with PEG 4600			
Molar ratio [PEG]/[DVS] = 1/33; molar ratio [PEG]/[cellulose] = 96/1			
Reagent	grams	mmoles	% by weight
Water	150	8330+280 ¹	91.80
CMCNa	2.25	3.21 * 10 ⁻³	1.38
HEC	0.75	3.00 * 10 ⁻³	0.46
KOH, 1M in water	5.28	KOH: 5.00	3.23
DVS	2.35	19.9	1.44
PEG 400	2.76	0.600	1.69

¹ the water of the KOH solution

Table 10

Hydrogel synthesis mixture with PEG 10,000			
Molar ratio [PEG]/[DVS] = 1/30; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
Water	150	8330+280 ¹	93.97
CMCNa	2.25	3.21 * 10 ⁻³	1.41
HEC	0.75	3.00 * 10 ⁻³	0.47
KOH, 1M in water	5.28	KOH: 5.00	3.31

¹ the water of the KOH solution

Table 10 (continued)

Hydrogel synthesis mixture with PEG 10,000 Molar ratio [PEG]/[DVS] = 1/30; molar ratio [PEG]/[cellulose] = 16/1			
Reagent	grams	mmoles	% by weight
DVS	0.35	2.96	0.22
PEG 400	1.00	0.100	0.63

10

Claims

1. A superabsorbent polysaccharide obtained by crosslinking a polysaccharide or derivative thereof with at least 1 % by weight of a flexible spacer having a chain length of at least 9 chain atoms and having terminal activated coupling groups.

15

2. A superabsorbent polysaccharide according to claim 1, in which said flexible spacer comprises a polyalkyleneglycol.

20

3. A superabsorbent polysaccharide according to claim 2, in which said polyalkyleneglycol has a molecular weight from about 400 to 10,000.

4. A superabsorbent polysaccharide according to claim 2 or 3, in which said polyalkyleneglycol is polyethyleneglycol.

25

5. A superabsorbent polysaccharide according to any one of claims 1-4, in which said coupling groups comprise vinyl sulphone groups.

30

6. A superabsorbent polysaccharide according to any one of claims 1-5, in which 10-67 % by weight of the flexible spacer, with respect to the polysaccharide, has been used.

35

7. A superabsorbent polysaccharide according to any one of claims 1-6, in which said polysaccharide has a molecular weight, before crosslinking, of between 100,000 and 1,500,000, preferably between 250,000 and 1,000,000.

40

8. A superabsorbent polysaccharide according to any one of claims 1-7, which has the form of a film having a thickness of between 10 and 500 µm.

9. A process for producing a superabsorbent polysaccharide according to any one of the preceding claims, comprising reacting a polyalkyleneglycol with at least two equivalents of a reagent containing one or more activated double bonds, such as divinyl sulphone, and reacting the polyalkyleneglycol having the double bonds thus obtained with a polysaccharide in the presence of a catalyst.

45

10. A hygiene product containing a superabsorbent polysaccharide film according to claim 8 or produced according to the process of claim 9.

50

55



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number
EP 00 20 0965

DOCUMENTS CONSIDERED TO BE RELEVANT			CLASSIFICATION OF THE APPLICATION (Int.Cl.7)						
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim							
A	WO 97 18890 A (STOCKHAUSEN GMBH) 29 May 1997 (1997-05-29) * abstract * * page 6, line 23 - line 36 * * page 10, line 16 - line 20 * * page 11, line 29 - page 12, line 2 * * claims-1,9,22-*	1-4,6-10	C08B37/00 C08B11/20 A61L15/28 A61L15/00						
A	US 5 414 135 A (SNOW ET AL.) 9 May 1995 (1995-05-09) * column 5, line 10 - line 26 *	9							
			TECHNICAL FIELDS SEARCHED (Int.Cl.7)						
			A61L						
<p>The present search report has been drawn up for all claims</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <tr> <td style="width: 33%;">Place of search</td> <td style="width: 33%;">Date of completion of the search</td> <td style="width: 34%;">Examiner</td> </tr> <tr> <td>THE HAGUE</td> <td>9 August 2000</td> <td>Mazet, J-F</td> </tr> </table> <p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons S : member of the same patent family, corresponding document</p>				Place of search	Date of completion of the search	Examiner	THE HAGUE	9 August 2000	Mazet, J-F
Place of search	Date of completion of the search	Examiner							
THE HAGUE	9 August 2000	Mazet, J-F							

EP 1 134 232 A1

**ANNEX TO THE EUROPEAN SEARCH REPORT
ON EUROPEAN PATENT APPLICATION NO.**

EP 00 20 0965

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report. The members are as contained in the European Patent Office EDP file on The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

09-08-2000

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO 9718890	A 29-05-1997	DE 19543368	A 22-05-1997	
		AT 185982	T 15-11-1999	
		CA 2235704	A 29-05-1997	
		CN 1207690	A 10-02-1999	
		CZ 9801505	A 11-11-1998	
		DE 59603516	D 02-12-1999	
		EP 0873188	A 28-10-1998	
		ES 2139394	T 01-02-2000	
US 5414135	A 09-05-1995	NONE		

8548

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82